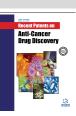
Recent Patents on Anti-Cancer Drug Discovery

#### **COMMENTARY**



# High-throughput Strategy Accelerates the Progress of Marine Anticancer Peptide Drug Development



Peng Lyu and Hang F. Kwok\*

Cancer Centre, Faculty of Health Sciences, University of Macau, University Avenue, Taipa, Macau, P.R. China

### 1. INTRODUCTION

With regard to the increasing requirement of novel anticancer drug, naturally occurring peptides/venom-based peptides are gaining much more attention. These natural peptides have demonstrated the convincing potential due to their high selectivity and diverse bio-functions in cancer cells. According to the literatures, more than 7000 therapeutic peptides have been successfully identified from different natural sources, including amphibians, snakes, scorpions, insects and marine cultures [1-3]. As of 2017, 484 therapeutic peptides are reported to be actively evaluated in different preclinical/clinical trial stages. Among these, 260 peptides have been tested in human clinical trials and more than 60 therapeutic peptides are approved by the FDA [4-6]. Although, the number of peptide therapies in the drug market is relatively small, the development of peptide-based therapies is reported to be steadily accelerating, where approximately 20 therapeutics peptides entered clinical trials per year in the past decade [7].

Marine sources itself have been receiving much more attention as a therapeutic peptide pool for the pharmaceutical industry in the last few years. According to the review article by Zheng and his colleagues [8], more than fifty marinebased anticancer peptide drugs have been well studied and patented since 2011. Several marine-based peptides/peptidomimetics have been clinically used to treat cancer [9]. In 2011, Adcetris (Brentuximab Vedotin), which is a marine peptide-based drug was approved by the FDA for Hodgkin lymphoma and ALCL (a type of T cell non-Hodgkin lymphoma) treatment [10]. On top of that, other marine peptidebased drugs such as Plitidepsin [11, 12], HTI-286 [13], Kahalalide F [14, 15], and Elisidepsin [16] are currently being assessed in different clinical trial phases for the treatment of various cancer types [17]. All these examples revealed that marine derived peptides have been proved to be a prolific source of anticancer drug discovery.

However, there are two major impediments that delay the development of marine source peptide-based drug discovery. The first challenge is that most of the marine source are very difficult to collect due to their small physical size and marine environment. Moreover, the application of 'classical' peptide

purification and identification techniques requires quite a large quantity of marine source to begin with. Consequently, only around 8% of marine peptides were identified and further studied [18]. Thus, a high-throughput platform combining transcriptome and proteome will be a more efficient and effective approach to accelerate the process of marine derived peptide discovery [19].

Another challenge is that the current technologies restrict a thorough mechanistic investigation of the anticancer peptide drugs. The traditional mechanism studies are based on certain targets and phenotype changes, which are also known as target-centric or phenotypic mechanism study. These traditional mechanism studies approach has been developed in the last decade, and several targets/pathways for anticancer peptides, such as the extrinsic (death receptor-mediated) pathway and the intrinsic (mitochondrial) pathway which induced apoptosis, p38 MAPK/ JNK pathway, tubulinmicrotubule disruption and PI3K/Akt pathway, have been verified and well-studied [8, 20]. However, as reported by Zheng and his colleagues [8], there is still only a small number of marine-derived anticancer peptides that are being intensively studied nowadays. More than 80% of the therapeutic peptide mechanisms still remain unclear even though these peptides are reported to be potential anti-cancer agents [8]. Insufficient mechanistic study always created a further delay in therapeutic development of marine-derived peptide into clinical stage. This predicament highlights the importance of a novel method which could allow a comprehensive and rapid mechanism study of therapeutic peptides, which is definitely essential.

# 2. HIGH-THROUGHPUT MARINE VENOM-DERIVED PEPTIDE DISCOVERY APPROACH

Traditional biochemistry, peptide identification, and purification processes such as microfiltration, gel filtration, reverse-phase HPLC, and ultracentrifugation are commonly utilized [21]. A large amount of crude venom/ marine source is required to unravel the complex peptide compounds in a single venom. In recent years, the dramatic development of mass spectrometry [22-24] and next-generation sequencing [25, 26] allowed high-throughput identification of venom peptides. The high resolution mass spectrometry approaches allowed the characterization of thousands of different peptides with a very small amount of crude venom/marine source [27]. Liquid Chromatography-Mass Spectrometry

<sup>\*</sup>Address correspondence to this author at the Faculty of Health Sciences, University of Macau, University Avenue, Taipa, Macau, P.R. China; E-mail: hfkwok@umac.mo

(LC-MS) is able to separate and analyze the venom mixtures in a single run. Typically, marine venoms were first lysed into peptide fragments. After the desalting process, the sample was injected into the high resolution LC-MS system. Large-scale peptide fragment sequence data were then obtained and analyzed by proteomics assemble software for de novo assemble [28-33]. In addition, transcriptome data can be obtained by next-generation sequencing techniques such as RNA-seq [19]. Through matching the proteomics and transcriptome data, a systematic overview of venom peptide library is provided beyond the efficiency of classical venom peptide identification approaches. This approach provides integrated, high resolution and rapid proteomics analysis of venoms. This cutting edge approach can accelerate the development of potential marine-derived therapeutic peptide discovery.

# 3. PATHWAY-CENTRIC ANTICANCER MECHA-NISM STUDY APPROACH

Most of the anticancer peptides induced multi-target and complex changes within the cancer cells. It is challenging to systematically identify their mechanism of action with any traditional biochemistry technologies [34]. A systematic and accurate approach using next-generation sequencing technique has been proposed to study the most relevant gene(s) responses, which are able to provide evidence in the whole transcriptome alterations after the therapeutic peptide treatment. Subsequently, bioinformatics method would able to further enrich the dysregulated transcriptome genes into several pathways. This approach retains the advantages of phenotypic and target-centric approaches and allows more comprehensive studies of the mechanism of anticancer peptides. Currently, the development of Next-Generation Sequence (NGS) technique allows the high-throughput and large-scale applications in drug discovery and clinical studies [35]. The broad utility of this approach will utmost enhance the efficiency and accuracy of marine-derived anticancer peptide mechanism study.

## 4. THE FUTURE OF MARINE ANTICANCER PEP-TIDE THERAPY

The rapid development of marine-derived anticancer peptide therapy enriches the anticancer drug pool, however, at the same time, exposes the shortcomings of traditional peptide identification and mechanism study methods. In the past years, the development of next-generation sequence and high-resolution mass-spectrometry techniques provided large-scale and high-throughput marine-derived peptide identification and comprehensive mechanism study. The robustness and broad utility of these novel methods have been proved [19, 36-38], and are capable to accelerate the discovery and development of marine anticancer peptide therapy.

#### **ACKNOWLEDGEMENTS**

This study was supported by the Science and Technology Development Fund of Macau SAR (FDCT) [019/2017/A1] to HFK. PL was in receipt of Postdoctoral Fellowship from the FDCT and the Faculty of Health Science University of Macau.

#### REFERENCES

- Fosgerau K, Hoffmann T. Peptide therapeutics: Current status and future directions. Drug Discov Today 2015; 20(1): 122-8.
- Padhi A, Sengupta M, Sengupta S, Roehm KH, Sonawane A. An-[2] timicrobial peptides and proteins in mycobacterial therapy: Current status and future prospects. Tuberculosis (Edinb) 2014; 94(4): 363-
- Buchwald H, Dorman RB, Rasmus NF, Michalek VN, Landvik NM, Ikramuddin S. Effects on GLP-1, PYY, and leptin by direct stimulation of terminal ileum and cecum in humans: Implications for ileal transposition. Surg Obes Relat Dis 2014; 10(5): 780-6.
- Lau JL, Dunn MK. Therapeutic peptides: Historical perspectives, [4] current development trends, and future directions. Bioorg Med Chem 2018; 26(10): 2700-7.
- [5] Vlieghe P, Lisowski V, Martinez J, Khrestchatisky M. Synthetic therapeutic peptides: Science and market. Drug Discov Today 2010; 15(1-2): 40-56.
- [6] Otvos L, Jr., Wade JD. Current challenges in peptide-based drug discovery. Front Chem 2014; 2: 62.
- Lax R. The future of peptide development in the pharmaceutical [7] industry. PharManufacturing: The Int Pep Rev 2010; 2: 10-5.
- [8] Zheng L, Xu Y, Lin X, Yuan Z, Liu M, Cao S, et al. Recent progress of marine polypeptides as anticancer agents. Recent Pat Anti-Cancer Drug Discov 2018; 13(4): 445-54.
- [9] Sable R, Parajuli P, Jois S. Peptides, peptidomimetics, and polypeptides from marine sources: A wealth of natural sources for pharmaceutical applications. Mar Drugs 2017; 15(4): doi: 10.3390/md15040124.
- Senter PD, Sievers EL. The discovery and development of brentuximab vedotin for use in relapsed Hodgkin lymphoma and systemic anaplastic large cell lymphoma. Nature Biotech 2012; 30(7): 631-8.
- Spicka I, Ocio EM, Oakervee HE, Greil R, Banh RH, Catley L, et al. Randomized Phase III study (ADMYRE) of plitidepsin in combination with dexamethasone vs. dexamethasone alone in patients with relapsed/refractory multiple myeloma. Am Soc Hematology; 2017; 89-96.
- Leisch M, Egle A, Greil R. Plitidepsin: A potential new treatment for relapsed/refractory multiple myeloma. Future Oncol 2018. doi: 10.2217/fon-2018-0492.
- [13] Loganzo F, Discafani CM, Annable T, Beyer C, Musto S, Hari M, et al. HTI-286, a synthetic analogue of the tripeptide hemiasterlin, is a potent antimicrotubule agent that circumvents P-glycoproteinmediated resistance in vitro and in vivo. Cancer Res 2003; 63(8):
- [14] Rademaker-Lakhai JM, Horenblas S, Meinhardt W, Stokvis E, de Reijke TM, Jimeno JM, et al. Phase I clinical and pharmacokinetic study of kahalalide F in patients with advanced androgen refractory prostate cancer. Clin Cancer Res 2005; 11(5): 1854-62.
- Martin-Algarra S, Espinosa E, Rubio J, Lopez Lopez JJ, Manzano JL, Carrion LA, et al. Phase II study of weekly Kahalalide F in patients with advanced malignant melanoma. Eur J Cancer 2009; 45(5): 732-5.
- [16] Goel S, Viteri S, Morán T, Coronado C, Dios JLI, Miguel-Lillo B, et al. Phase I, dose-escalating study of elisidepsin (Irvalec®), a plasma membrane-disrupting marine antitumor agent, in combination with erlotinib in patients with advanced malignant solid tumors. Invest New Drugs 2016; 34(1): 75-83.
- [17] Cheung RC, Ng TB, Wong JH. Marine peptides: Bioactivities and applications. Mar Drugs 2015; 13(7): 4006-43.
- [18] Hu Y, Chen J, Hu G, Yu J, Zhu X, Lin Y, et al. Statistical research on the bioactivity of new marine natural products discovered during the 28 years from 1985 to 2012. Mar Drugs 2015; 13(1): 202-
- [19] Li B, Lyu P, Xi X, Ge L, Mahadevappa R, Shaw C, et al. Triggering of cancer cell cycle arrest by a novel scorpion venom-derived peptide-gonearrestide. J Cell Mol Med 2018; 22(9): 4460-73.
- [20] Ma R, Mahadevappa R, Kwok HF. Venom-based peptide therapy: Insights into anti-cancer mechanism. Oncotarget 2017; 8(59): 100908-30.

- [21] Lemes AC, Sala L, Ores Jda C, Braga AR, Egea MB, Fernandes KF. A Review of the Latest Advances in Encrypted Bioactive Peptides from Protein-Rich Waste. Int J Mol Sci 2016; 17(6).
- [22] Domon B, Aebersold R. Mass spectrometry and protein analysis. Science 2006; 312(5771): 212-7.
- [23] Graves PR, Haystead TA. Molecular biologist's guide to proteomics. Micro Mol Bio Rev 2002; 66(1): 39-63.
- [24] Jensen ON. Interpreting the protein language using proteomics. Nature reviews Mol Cell Bio 2006; 7(6): 391-8.
- [25] Mardis ER. Next-generation sequencing platforms. Annu Rev Anal Chem 2013; 6: 287-303.
- [26] Quail MA, Smith M, Coupland P, Otto TD, Harris SR, Connor TR, et al. A tale of three next generation sequencing platforms: Comparison of ion torrent, pacific biosciences and illumina MiSeq sequencers. BMC Genomics 2012; 13(1): 341.
- [27] Aebersold R, Mann M. Mass-spectrometric exploration of proteome structure and function. Nature 2016; 537(7620): 347-55.
- [28] Tran NH, Zhang X, Xin L, Shan B, Li M. De novo peptide sequencing by deep learning. Proc Natl Acad Sci USA 2017; 114(31): 8247-52.
- [29] Tran NH, Rahman MZ, He L, Xin L, Shan B, Li M. Complete de novo assembly of monoclonal antibody sequences. Sci Rep 2016; 6: 31730-7.
- [30] Zhang J, Xin L, Shan B, Chen W, Xie M, Yuen D, et al. PEAKS DB: De novo sequencing assisted database search for sensitive and accurate peptide identification. Mol Cell Proteomics 2012; 11(4): M111.010587.

- [31] Dutertre S, Jin A-h, Kaas Q, Jones A, Alewood PF, Lewis RJ. Deep venomics reveals the mechanism for expanded peptide diversity in cone snail venom. Mol Cell Proteomics. 2013; 12(2): 312-29.
- [32] Himaya S, Jin A-H, Dutertre Sb, Giacomotto J, Mohialdeen H, Vetter I, *et al.* Comparative venomics reveals the complex prey capture strategy of the piscivorous cone snail Conus catus. J Proteome Res 2015; 14(10): 4372-81.
- [33] Jin A-h, Dutertre S, Kaas Q, Lavergne V, Kubala P, Lewis RJ, et al. Transcriptomic messiness in the venom duct of Conus miles contributes to conotoxin diversity. Mol Cell Proteomics 2013: MCP. M113.030353.
- [34] Flordellis CS, Manolis AS, Paris H, Karabinis A. Rethinking target discovery in polygenic diseases. Curr Top Med Chem 2006; 6(16): 1791-8
- [35] Li H, Zhou H, Wang D, Qiu J, Zhou Y, Li X, et al. Versatile pathway-centric approach based on high-throughput sequencing to anticancer drug discovery. Proc Natl Acad Sci USA 2012; 109(12): 4609-14.
- [36] Robinson SD, Undheim EA, Ueberheide B, King GF. Venom peptides as therapeutics: Advances, challenges and the future of venom-peptide discovery. Expert Rev Proteomics 2017; 14(10): 931-9.
- [37] Prashanth JR, Lewis RJ. An efficient transcriptome analysis pipeline to accelerate venom peptide discovery and characterisation. Toxicon 2015; 107: 282-9.
- [38] Xie B, Huang Y, Baumann K, Fry BG, Shi Q. From marine venoms to drugs: Efficiently supported by a combination of transcriptomics and proteomics. Marine Drugs 2017; 15(4): 103-10.